

	UNIVERSITY OF CAMBRIDGE INTE General Certificate of Education Advanced Subsidiary Level and Advar	
CANDIDATE NAME		
CENTER NUMBER		CANDIDATE NUMBER
BIOLOGY (US)		9184/3
Advanced Pract	tical Skills 1	May/June 20
0		2 hou
	wer on the Question Paper.	
Additional Mate	rials: As listed in the Confidential Inst	uctions.

READ THESE INSTRUCTIONS FIRST

Write your Center number, candidate number and name on all the work you hand in. Write in dark blue or black ink.

You may use a pencil for any diagrams, graphs or rough working. Do **not** use red ink, staples, paper clips, highlighters, glue or correction fluid. DO NOT WRITE IN ANY BARCODES.

Answer all questions. Electronic calculators may be used. You may lose marks if you do not show your working or if you do not use appropriate units.

At the end of the examination, fasten all your work securely together.

The number of marks is given in brackets [] at the end of each question or part question.

For Examiner's Use		
1		
2		
Total		

This document consists of 15 printed pages and 1 blank page.



You are reminded that you have only one hour for each question in the practical examination

You should:

- read carefully through the whole of Question 1 and Question 2 •
- www.PapaCambridge.com then plan your use of the time to make sure that you finish all the work that you would like to do.

You will gain marks for recording your results according to the instructions.

1 Glucose solutions change the color of pink potassium manganate(VII) solution, PM.

Fig. 1.1 shows the color change from pink to the colorless end-point.



Fig. 1.1

The rate of the color change depends on the concentration of the glucose solution. The greater the concentration of glucose solution the faster the end-point is reached.

You are required to:

- make different concentrations of glucose solution
- find, for each glucose solution, the time taken for **PM** to change to colorless
- estimate the unknown concentrations of the glucose solutions, **U1** and **U2**.

You are provided with:

ure provided wi	3 th:		volume/cm³ 100 200	Papac
labeled	contents	hazard	volume/cm ³	
G	20% glucose solution	none	100	
W	distilled water	none	200	
S	sulfuric acid	harmful	40	
РМ	potassium manganate(VII) solution	harmful	20	
U1	glucose solution	none	20	
U2	glucose solution	none	20]

Sulfuric acid and potassium manganate(VII) solution are harmful. If any comes into contact with your skin, wash immediately under cold water. It is recommended that you wear safety glasses.

Proceed as follows:

- Using the 20% glucose solution, G, as a starting concentration you are required to 1. make up 20 cm³ of each of four different concentrations of glucose solutions, 6%, 8%, 10%, 12%.
- (a) (i) Complete Table 1.1 to show how you will make the four glucose solutions 6%, 8%, 10% and 20%.

volume of 20% glucose solution/cm ³	volume of distilled water/cm ³	final percentage concentration of glucose
		6
		8
		10
12	8	12

Та	b	e	1	.1	
		-			

[2]

- 2. Make all the glucose solutions as in Table 1.1, in the containers provided.
- 3. Put 10 cm³ of each glucose solution into four separate test-tubes.
- Using the syringe labeled **S**, put 5 cm^3 of **S** into each test-tube. Insert the rubber 4. stopper and, with your finger holding the rubber stopper in place, gently mix the solution in each test-tube. Do **not** turn the test-tube upside-down.

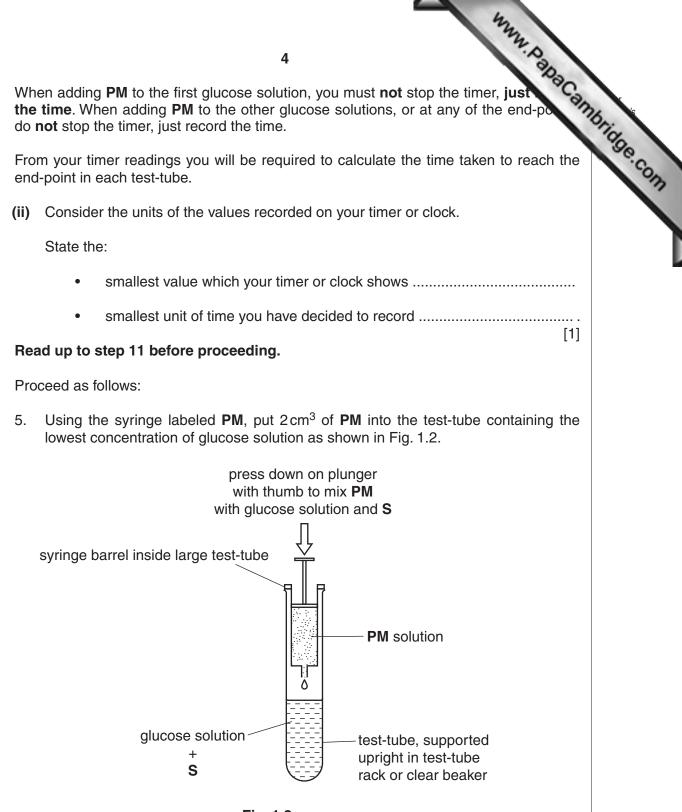


Fig. 1.2

- 6. Start timing and record the start time from your timer on Fig. 1.3 on page 5.
- 7. Immediately, put 2 cm³ of **PM** into the test-tube containing next highest concentration of glucose solution.
- 8. Record start time from your timer on Fig. 1.3 on page 5.
- 9. Immediately, repeat steps 7 and 8 for the remaining concentrations of glucose solution.

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5

10. Observe the four test-tubes and record the time on Fig. 1.3 when each end reached.

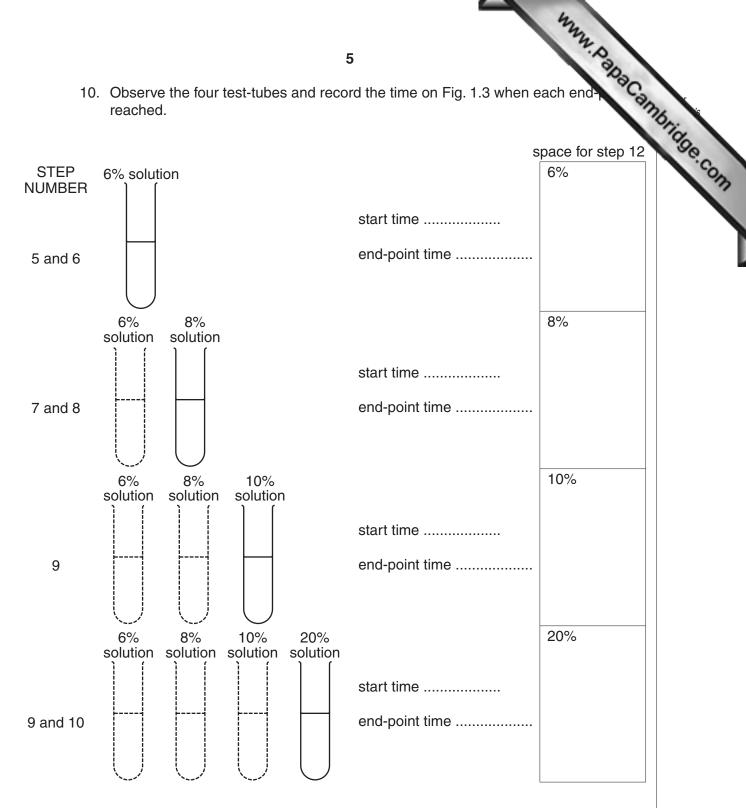


Fig. 1.3

www.papaCambridge.com 6 You are required to estimate the glucose concentration of solutions, U1 and U the same procedure. State one variable, which you will standardize when setting up the test-tubes to find (iii) the end-points for U1 and U2.[1] (iv) Describe how you will standardize this variable.[1] 11. Use the same procedure to obtain the end-points for the solutions, U1 and U2 and record your times on Fig. 1.4. space for step 12 **U1 U1** start time end-point time U2 U1 U2 start time end-point time

Fig. 1.4

[3]

www.papacambridge.com Depending on the timer or clock you have used, you may find the following example. helpful so that you can process your results for (v) and for Step 12 to find the time ta to reach the end-point.

Example 1: using stop-clock or stopwatch

start time end-point time	minutes:seconds 1:24 2:55	= 84 seconds = 175 seconds
time	taken to reach end-point	= 91 seconds

Example 2: using clock times

start time end-point time	hours:minutes:seconds 9:10:00 9:11:31	difference in time 1 minute 31 seconds		
time taken to reach end-point = 91 seconds				

(v) Using your results from Fig. 1.3, complete Table 1.2 to show the calculation to find the time taken for 6% glucose solution to reach the end-point.



6% solution start time 6% solution end-point time		
ti	me taken to reach end-point	=

12. Use the space on Fig. 1.3 and Fig. 1.4 for processing your readings to find the time taken to reach the end-point for the other glucose solutions, and for **U1** and **U2**.

www.papaCambridge.com (vi) Using only these processed results, prepare the space below to record the taken to reach the end-point for all six solutions.

Glucose solutions may be used for different purposes, for example:

Glucose tolerance test solutions, containing 25% glucose.

Sports drink solutions, containing 8% glucose.

Oral Rehydration solutions, containing 2% glucose.

(b) Suggest which of the above solutions is U2.

.....[1]

12	
9	
Identify one significant source of error in your investigation.	
	Orio
[1]	
Describe two modifications to this investigation which would improve the confidence in your results.	
[2]	
[Total: 17]	
	Identify one significant source of error in your investigation. [1] Describe two modifications to this investigation which would improve the confidence in your results. [2]

Question 2 starts on page 10

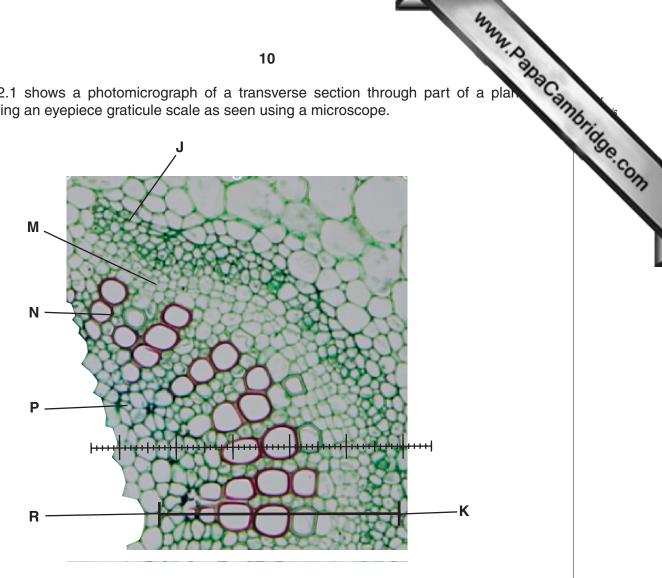


Fig. 2.1

An eyepiece graticule scale can be used to measure the layers of tissues and to help draw a plan diagram with the correct shape and proportions of the tissues, without needing to calibrate the eyepiece graticule scale.

(a) (i) The length of the vascular bundle (from K to R) in Fig. 2.1 was measured using the eyepiece graticule scale and recorded in Table 2.1.

Table 2.1

layer	J	М	Ν	Р	length from K to R
number of eyepiece graticule scale divisions					43

Complete Table 2.1 by finding the thickness of the different layers L, M, N and P, labeled in Fig. 2.1, using the line between R and K and the eyepiece graticule scale. [2]

The length (from K to R) of the vascular bundle in eyepiece graticule division used to make a scale drawing of the outline of the vascular bundle as show Fig. 2.2.

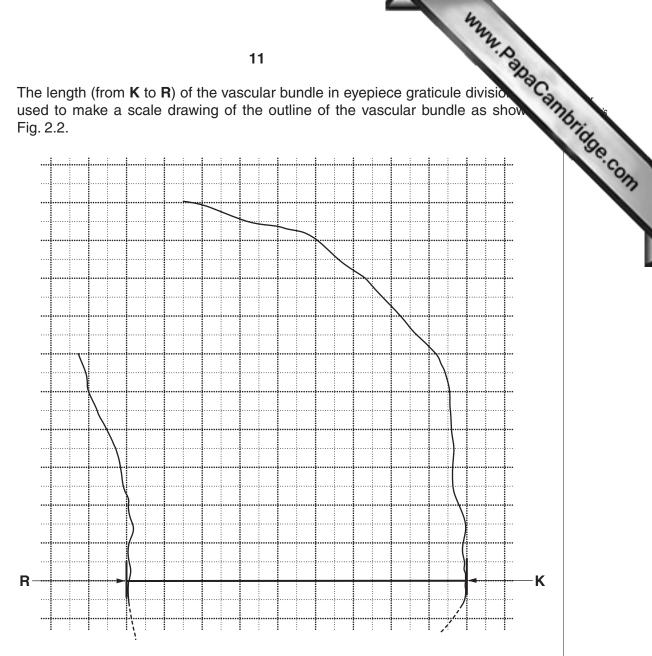


Fig. 2.2

- (ii) Complete the plan diagram of this part of the vascular bundle to show the proportions and shape of each of the tissues. Use the values in Table 2.1 to help you. [3]
- Using Fig. 2.2, count the total number of 1 cm by 1 cm squares occupied by the (iii) vascular bundle and count the total number of 1 cm by 1 cm squares occupied by the xylem tissue.

Count any 'half square' or 'more than half' as one square.

State the ratio of the area occupied by the vascular bundle to that of the xylem tissue.

You will lose marks if you do not show all the steps in finding the ratio, including indicating counted squares on Fig. 2.2.

ratio

L1 is a slide of a transverse section through the same plant stem as in Fig. 2.1. This plant grows mainly in Europe and Asia.

www.papaCambridge.com This stem shows a stained tissue, close to the epidermis, in the four corners of the stem. Near to the center of the stem is a different tissue.

- (b) Make a large drawing of one group of three whole, adjacent (touching) cells
 - from the tissue in one corner, as observed on the specimen on L1.

Make a large drawing of one group of three whole, adjacent cells

from the tissue near to the center of the stem, as observed on the specimen on L1.

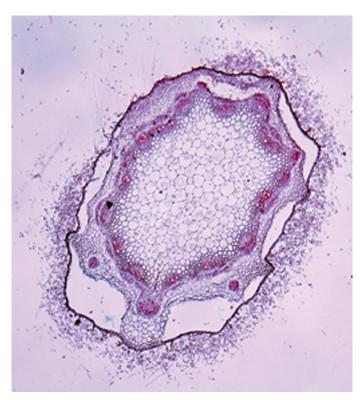
The drawings should show any difference in size (linear magnification) observed between each group of cells.

On your drawing, use a ruled label line and label to show one cell wall.

cells from the tissue in one corner

cells from the tissue near to the center of the stem

www.papaCambridge.com Fig. 2.3 is a photomicrograph of a transverse section through a stem of a different species.





- (c) Prepare the space below so that it is suitable for you to record observable differences between the specimens on slide L1 and in Fig. 2.3 to include:
 - the vascular tissue
 - at least two other tissues.

Some scientists carried out an investigation into the uptake of glucose by five different of plant tissues during the course of 25 minutes.

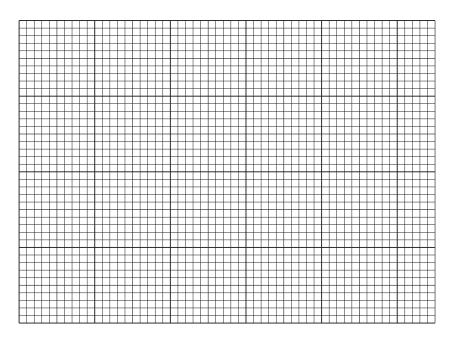
A piece of each type of plant tissue was placed in a solution of glucose.

www.papacambridge.com The starting concentration of this glucose solution was 0.8 arbitrary units which was lower than the concentration inside the plant cells in each tissue.

The results after 25 minutes are shown in Table 2.2.

type of plant tissue	concentration of glucose in the cells/arbitrary units
А	2.0
В	6.5
С	4.2
D	5.6
E	3.2

(d) (i) Plot a chart of the data shown in Table 2.2.



[4]

	15 Describe and explain the results shown in the chart you have drawn.	
	15	
(ii)	Describe and explain the results shown in the chart you have drawn.	
		orido
		°.con
		12
	[3]	
	[Total: 23]	



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